

## 201. Sponge-Derived Polyunsaturated C<sub>16</sub> Di- and Tribromocarboxylic Acids

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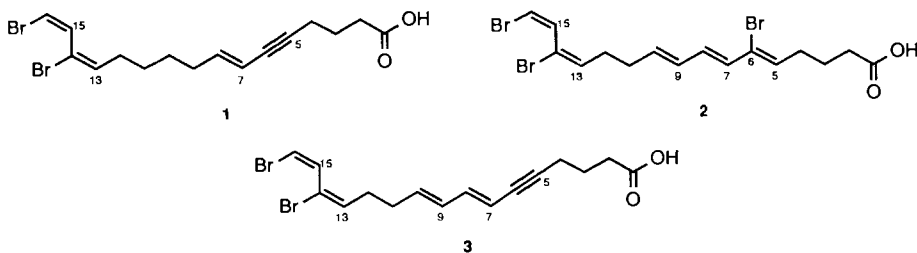
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One previously described and two new bromo-substituted polyunsaturated C<sub>16</sub> fatty acids were isolated from an Indonesian sponge, *Oceanapia* sp. Their common structural feature is an (13*E*,15*Z*)-14,16-dibromodiene terminus. They differ in their C(5) to C(10) portions in unsaturation and halogenation. All three compounds are unstable oils. The mixture exhibits mild cytotoxicity against KB cells.

**Introduction.** – Sponges of the genera *Xestospongia* and *Petrosia* (order Petrosida, family Petrosiidae) yielded a number of brominated polyunsaturated fatty acids [1–7]. A majority of them (21) are C<sub>18</sub> acids, but six C<sub>16</sub> and one C<sub>9</sub> acid are also represented. The unsaturation, except for the C<sub>9</sub> compound, includes olefinic and acetylenic functions. The first of these metabolites, (7*E*,13*E*,15*Z*)-14,16-dibromohexadeca-7,13,15-trien-5-ynoic acid (**1**), was reported by Schmitz and Gopichand [1] as a CNS-active constituent of the sponge *Xestospongia muta*. Its unique structural feature is its (13*E*,15*Z*)-14,16-dibromodiene terminus.

We now report the isolation of **1** and of two new compounds, (5*Z*,7*E*,9*E*,13*E*,15*Z*)-6,14,16-tribromohexadeca-5,7,9,13,15-pentaenoic acid (**2**) and (7*E*,9*E*,13*E*,15*Z*)-14,16-dibromohexadeca-7,9,13,15-tetraen-5-ynoic acid (**3**) from a sponge, *Oceanapia* sp. (order Petrosida, family Oceanapiidae), collected in October, 1992, off Manado, Sulawesi, Indonesia. All three acids have a common dibromodiene terminus.



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**Results and Discussion.** – The freeze-dried sponge was extracted with EtOH. The residue after solvent removal was partitioned between  $\text{CH}_2\text{Cl}_2$  and  $\text{H}_2\text{O}$  and yielded 2.64 g of lipid extract. A portion of this (420 mg) was subjected to high-speed countercurrent chromatography, followed by reversed-phase HPLC, which resulted in the isolation of three unstable yellow oils, **1** (7.9 mg), **2** (7.5 mg), and **3** (26.9 mg). The structures of these compounds were established using  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR and mass spectroscopy and, in the case of **1**, by comparison with known data [1].

The  $^{13}\text{C}$ -NMR spectrum of **1** has 16 signals. The presence of an acid ( $\delta$  178.7), an acetylenic ( $\delta$  87.1 and 80.1), and 3 olefinic moieties ( $\delta$  143.2, 134.5, 131.0, 113.6, 112.4, 109.9) is readily apparent. The  $^1\text{H}$ -NMR spectrum of **1** shows the presence of 1 disubstituted (*E*)-olefin at  $\delta$  5.44 (*d*,  $J = 14.4$  Hz) and 6.03 (*dt*,  $J = 15.3$ , 7.8 Hz), 1 disubstituted (*Z*)-olefin at  $\delta$  6.68 (*dd*,  $J = 7.7$  Hz) and 6.38 (*d*,  $J = 7.8$  Hz), and a trisubstituted olefin at  $\delta$  6.06 (*t*,  $J = 7.7$  Hz; see *Table*). These NMR data indicate that compound **1** is a  $\text{C}_{16}$  polyunsaturated acetylenic acid. A comparison with published  $^1\text{H}$ -NMR data establishes the identity of **1** and (*7E*, *13E*, *15Z*)-14,16-dibromohexadeca-7,13,15-trien-5-ynoic acid [1].

Table.  $^1\text{H}$ -NMR Data of Compounds **1**, **2**, and **3**.  $\delta$  in ppm and  $J$  in Hz.

	<b>1</b>	<b>2</b>	<b>3</b>
$\text{CH}_2(2)$	2.50 ( <i>t</i> , $J = 7.5$ )	2.40 ( <i>t</i> , $J = 7.4$ )	2.50 ( <i>t</i> , $J = 7.4$ )
$\text{CH}_2(3)$	1.85 ( <i>quint.</i> , $J = 7.1$ )	1.79 ( <i>quint.</i> , $J = 7.4$ )	1.86 ( <i>quint.</i> , $J = 7.1$ )
$\text{CH}_2(4)$	2.38 ( <i>dt</i> , $J = 1.6$ , 6.8)	2.38 ( <i>q</i> , $J = 7.3$ ), 5.90 ( <i>t</i> , $J = 7.2$ )	2.42 ( <i>dt</i> , $J = 1.4$ , 6.6)
H–C(5)			
H–C(7)	5.44 ( <i>d</i> , $J = 14.4$ )	6.11 ( <i>d</i> , $J = 14.4$ )	5.49 ( <i>br. d</i> , $J = 15.6$ )
H–C(8)	6.03 ( <i>dt</i> , $J = 15.3$ , 7.8)	6.53 ( <i>dd</i> , $J = 10.5$ , 14.4)	6.47 ( <i>dd</i> , $J = 10.7$ , 15.5)
$\text{CH}_2(9)$ or H–C(9)	2.01 ( <i>q</i> , $J = 6.6$ )	6.14 ( <i>dd</i> , $J = 11.6$ , 14.6)	6.07 ( <i>dd</i> , $J = 11.7$ , 14.5)
$\text{CH}_2(10)$ or H–C(10)	1.40 ( <i>m</i> )	5.79 ( <i>dt</i> , $J = 15.3$ , 7.1)	5.70 ( <i>dt</i> , $J = 6.6$ , 15.3)
$\text{CH}_2(11)$	1.40 ( <i>m</i> )	2.25 ( <i>q</i> , $J = 7.1$ )	2.22 ( <i>q</i> , $J = 6.6$ )
$\text{CH}_2(12)$	2.07 ( <i>q</i> , $J = 6.6$ )	2.13 ( <i>q</i> , $J = 7.3$ )	2.12 ( <i>q</i> , $J = 7.1$ )
H–C(13)	6.06 ( <i>t</i> , $J = 7.7$ )	6.07 ( <i>dt</i> , $J = 1.2$ , 7.5)	6.05 ( <i>t</i> , $J = 6.3$ )
H–C(15)	6.68 ( <i>dd</i> , $J = 1.1$ , 7.7)	6.68 ( <i>dd</i> , $J = 1.1$ , 7.7)	6.68 ( <i>d</i> , $J = 7.5$ )
H–C(16)	6.38 ( <i>d</i> , $J = 7.8$ )	6.39 ( <i>d</i> , $J = 7.8$ )	6.39 ( <i>d</i> , $J = 7.5$ )

Compound **2** has the molecular formula  $\text{C}_{16}\text{H}_{21}\text{Br}_3\text{O}_2$ , as shown by the HR-EI-MS data. Some signals in the  $^1\text{H}$ -NMR spectrum of **2** are identical with those of **1** (*Table*). The lack of an acetylene moiety is indicated by the IR and  $^{13}\text{C}$ -NMR spectra. The  $^1\text{H}$ -NMR spectrum shows 8 olefinic-proton and 5 aliphatic-proton signals. Decoupling experiments connect  $\text{CH}_2(2)$  to  $\text{CH}_2(3)$  to  $\text{CH}_2(4)$  and the latter to H–C(5) ( $\delta$  5.90 (*t*,  $J = 7.2$  Hz)). Additional decoupling experiments reveal the H–C(7) to H–C(13) sequence which is readily extended to H–C(16) by comparison with the  $^1\text{H}$ -NMR data of **1**. Since the COOH group is unconjugated (IR:  $1700\text{ cm}^{-1}$ ;  $^{13}\text{C}$ -NMR: 178.5 ppm), it necessarily must be attached to C(2) and the third Br-atom to C(6). Coupling constants between H–C(7) and H–C(8), H–C(9) and H–C(10), H–C(15) and H–C(16) allow the assignment of the (*7E*, *9E*, *15Z*)-configuration. NOE's from H–C(5) to H–C(7) and from  $\text{CH}_2(12)$  to H–C(15) establish the (*5Z*, *13E*)-configuration<sup>2</sup>). *Schmitz* and *Gopichand* [1] did not assign the C(13)=C(14) geometry of compound **1**. But since the  $^1\text{H}$ -NMR chemical shift for H–C(13) of **2** is almost the same as for **1**, the latter could also have (*13Z*)-configuration.

HR-EI-MS of compound **3** leads to a molecular formula  $\text{C}_{16}\text{H}_{18}\text{Br}_2\text{O}_2$ , differing from **1** by  $\text{H}_2$ .  $^1\text{H}$ -NMR spectral comparison (*Table*) shows that **3** differs from **1** by an additional olefin double bond at C(9)=C(10).

Since compounds **1–3** are unstable when pure, cytotoxicity was tested on the mixture, which showed only weak cytotoxicity (2+ at  $10\text{ }\mu\text{g/ml}$ ) against KB cells. Compound **3** showed mild antimicrobial activity against Gram-positive bacteria.

<sup>2</sup>) An unusually long relaxation time of 10 s was required to observe an NOE between  $\text{CH}_2(12)$  and H–C(15).

## Experimental Part

*General.* Anal. TLC: pre-coated HP-TLC plates (silica gel 60 F254 and RP-18 F254s). Countercurrent chromatography: *Ito-Multi-Layer-Coil* separator-extractor. Reversed-phase HPLC: *YMC-Pack AQ-ODS*. UV:  $\lambda_{\max}$  ( $\epsilon$ ) in nm. IR: NaCl plates; in  $\text{cm}^{-1}$ . NMR:  $\delta$  in ppm rel. to  $\text{Me}_4\text{Si}$  ( $= 0$  ppm),  $J$  in Hz.

*Isolation.* A sponge was collected from Manado Bay, Sulawesi, Indonesia, on October 1, 1992, from a depth of 10–40 m. The sponge is a semi-spherical to thickly encrusting mass, with broad papillar oscular projections, firm in life and when preserved. The sponge is yellowish-orange to white, with a reddish tinge on light-exposed surfaces, and is beige in EtOH preservative. It is most closely comparable to *Oceanapia papula* DESQUEYROUX-FAUNDEZ 1987 (Porifera, Demospongiae, Petrosida, Oceanapiidae). A voucher specimen was deposited at the Harbor Branch Oceanographic Museum, Fort Pierce, Florida (Catalog No. 033: 890). A freeze-dried sponge (dry weight 104 g) was extracted with EtOH ( $4 \times 900$  ml). The solvent was evaporated and the resulting residue partitioned between  $\text{CH}_2\text{Cl}_2$  and  $\text{H}_2\text{O}$  to yield 2.64 g of nonpolar extract. A portion of the nonpolar extract (420 mg) was applied to a high-speed countercurrent chromatography (HS-CCC) coil with heptane/MeCN/ $\text{CH}_2\text{Cl}_2$  10:7:3 (lower mobile phase). The second fraction from HS-CCC separation was purified by reversed-phase HPLC (MeOH/ $\text{H}_2\text{O}$  85:15): **1** (7.9 mg), **2** (7.5 mg), and **3** (26.9 mg) as yellow oils.

(7E,13E,15Z)-14,16-Dibromohexadeca-7,13,15-trien-5-ynoic Acid (**1**).  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ ): Table.  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ ): 178.7 (s), 143.2 (d); 136.5 (d); 131.0 (d); 113.6 (s); 112.4 (d); 109.9 (d); 87.1 (s); 80.1 (s); 32.6 (2t); 30.9 (t); 28.2 (t); 28.0 (t); 23.6 (t); 18.7 (t). EI-MS: 402, 323, 321, 279, 263, 243, 183, 117, 91.

(5Z,7E,9E,13E,15Z)-6,14,16-Tribromohexadeca-5,7,9,13,15-pentaenoic Acid (**2**). UV (MeOH): 208 (4600), 252 (sh, 7300), 264 (11700), 272 (14600), 284 (11900). IR (neat): 3500–2500 (br.), 1700, 1590, 1430, 975.  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ ): Table.  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ ): 178.5 (s); 135.5 (d); 135.3 (d); 133.1 (d); 132.2 (d); 131.0 (d); 130.2 (d); 129.5 (d); 126.1 (d); 113.9 (s); 112.5 (d); 33.1 (t); 31.7 (t); 30.9 (t); 30.8 (t); 23.5 (t). EI-MS: 486, 484, 482, 177, 117, 80. HR-EI-MS: 481.911118 ( $\text{C}_{16}\text{H}_{21}\text{Br}_3\text{O}_2$ , calc. 481.90943).

(7E,9E,13E,15Z)-14,16-Dibromohexadeca-7,9,13,15-tetraen-5-ynoic Acid (**3**). UV (MeOH): 204 (15000), 256 (sh, 29000), 268 (37100), 278 (28700). IR (neat): 3500–2500 (br.), 3010, 2930, 2200, 1700, 1590, 980.  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ ): Table.  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ ): 179.5 (s); 140.8 (d); 135.4 (d); 135.0 (d); 130.9 (d); 130.7 (d); 114.0 (s); 112.5 (d); 109.9 (d); 90.8 (s); 80.8 (s); 32.8 (t); 31.6 (t); 30.7 (t); 23.7 (t); 19.0 (t). EI-MS: 404, 402, 400, 323, 321, 241, 177, 117, 91. HR-EI-MS: 399.967407 ( $\text{C}_{16}\text{H}_{18}\text{Br}_2\text{O}_2$ , 399.96754).

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